



A polymeric micellar drug delivery system developed through a design of Experiment approach improves pancreatic tumor accumulation of calcipotriol and paclitaxel

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1. Introduction

Despite intense efforts, pancreatic ductal adenocarcinoma (PDAC) remains a treatment-refractory malignancy with a 5-year survival rate of 6% (Hall et al., 2018; Mcguigan et al., 2018). A literature search shows the survival rate is quite diverse, ranging from 2% to 9% (Ilic and Ilic, 2016; Luo et al., 2013). Gemcitabine was shown to improve symptoms and extend survival in patients (Burris et al., 1997). However, since 1997, only two systemic therapies have shown marginal improvement in survival compared to gemcitabine monotherapy, but with serious side effects including febrile neutropenia, cardiac decompensation and sensory neuropathy which sometimes result in death (Vaccaro et al., 2011). The characteristic dense stromal cells in pancreatic tumors are known to impede drug delivery and contribute to chemoresistance.

A strategy that targets both tumor and stroma and improves drug delivery into these compartments has since been proposed as an alternative that could significantly improve survival outcomes in PDAC (Zhao et al., 2018a). Recently, it was shown that vitamin-D receptor-mediated stromal reprogramming could improve pancreatic cancer therapy (Sherman et al., 2014). Calcipotriol (Cal), a synthetic vitamin-D analogue has shown promise in its ability to de-activate cancer associated fibroblasts within the stromal milieu, effectively “breaking” the stromal barrier to enhance the delivery of chemotherapy into the tumor (Sherman et al., 2014). Even as the least hypercalcemia-inducing compound within the vitamin-D derivatives class, Cal remains toxic at high doses, limiting its potential clinical use. Cal is an agonist of the vitamin D receptor (VDR), a ubiquitously expressed receptor found in a variety of tissues and controls the activity of several genes. Following activation, the VDR triggers a cascade of reactions that result in modifications in transcriptional output of genes (Pike and Meyer, 2012). Therefore,

targeted activation of the VDR within the stroma is desirable to reduce off-target effects. Nanotechnology-based drug delivery systems have been proposed to offer the possibilities for efficient tumor targeting with diminished drug-related adverse events (Meng and Nel, 2018).

This study focused on developing a nano-polymeric micellar drug delivery system, loaded with Cal and PTX for the treatment of PDAC, using a Design of Experiment (DOE) approach. Two nanotechnology-based therapeutics, namely, Abraxane (albumin-bound PTX) and Onivyde (liposomal formulation of irinotecan), are currently available and prescribed for various types of cancer (Shi et al., 2017; Zhang et al., 2016). Conjugation of albumin to PTX improved its toxicity profile when compared to cremophor-based PTX formulation (Kim, 2017), and a combination regimen of albumin-bound PTX and gemcitabine outperformed gemcitabine monotherapy. Albumin-bound PTX is recognized to have some effects on stromal cells and modulates the tumor microenvironment. The improved therapeutic effect is largely attributed to this stromal-modulatory properties (Alvarez et al., 2013). However, these gains remain very modest. To this effect, our dual-payload drug delivery platform is designed to carry both a cytotoxic payload (PTX), and a bonafide stroma-modulating agent (Cal) for simultaneous targeting of stromal and tumor compartments.

We systematically optimized the delivery vehicle using the DOE approach to obtain a pre-specified micelle size and extend the blood circulation times of Cal and PTX. DOE is a mathematical modeling paradigm that involves developing relationships between process variables and response/output. Traditionally, at least in the area of drug development, this approach has been confined to optimizing process parameters (Gupta et al., 2015; Hejri et al., 2013; Zhang et al., 2013). Here, we extended its application to the selection of monomer units and degree of crosslinking in the synthesis of a polymeric drug delivery

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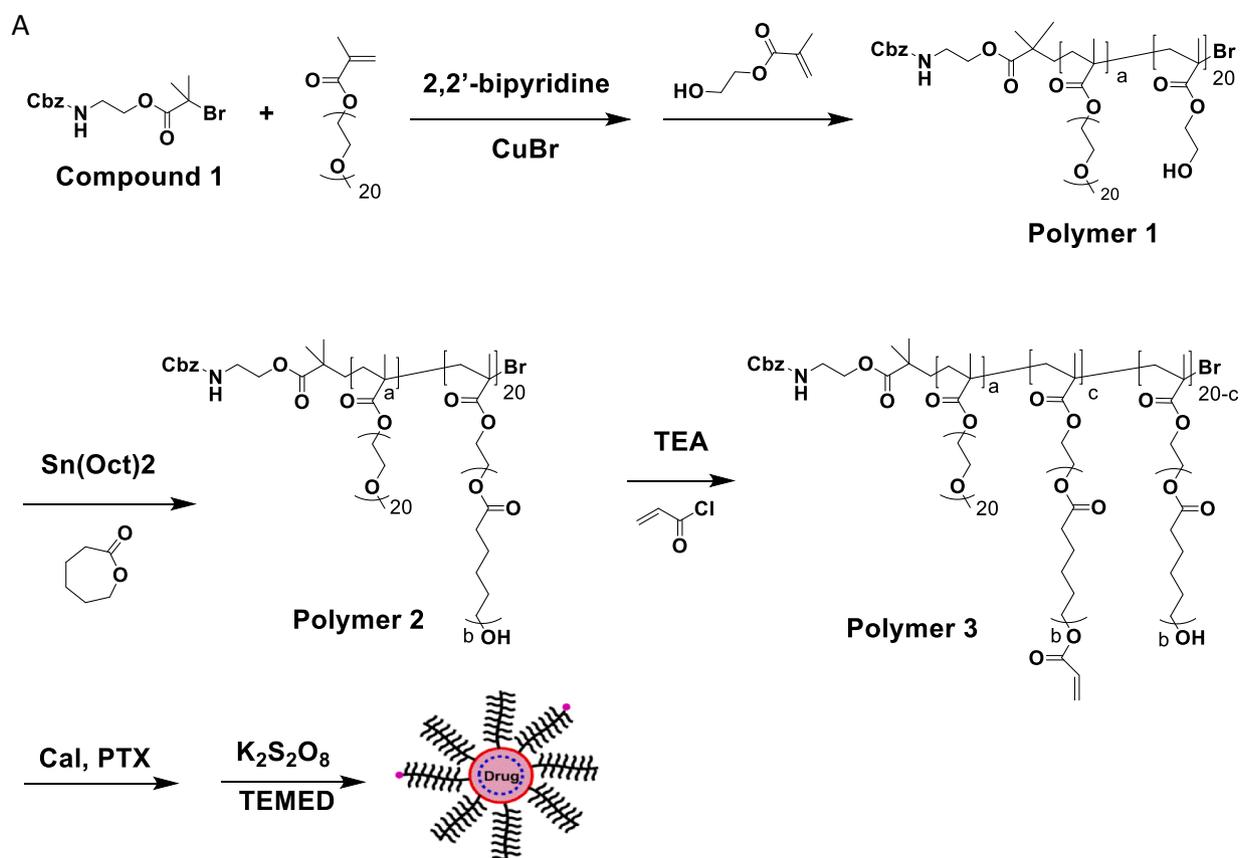
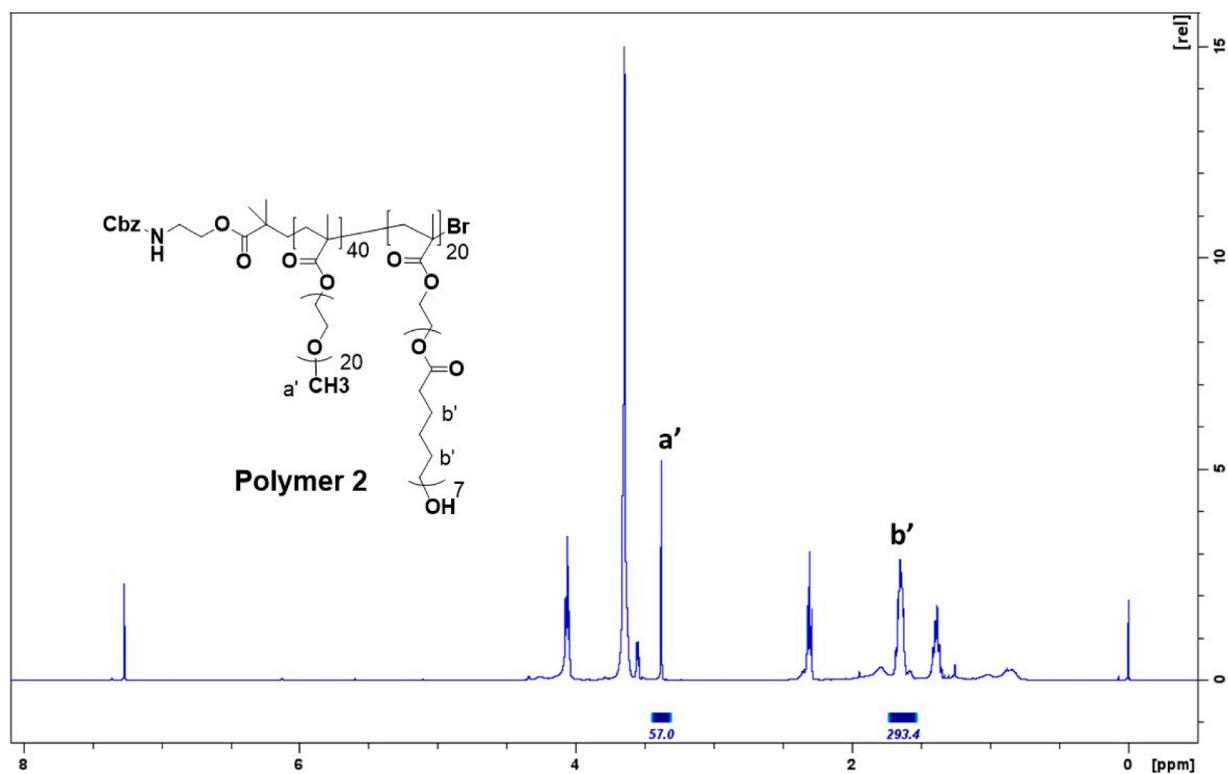
**B.**

Fig. 1. (A) Polymer synthesis and formulation of Cal and PTX loaded micelles (M-Cal/PTX). TEA, trimethylamine; TEMED, tetramethylethylenediamine. The values a, b, c were variables used in the central composite design (CCD). (B) ^1H NMR spectrum of poly(OEG-MA)₄₀-b-poly[HEMA-g-(ε-caprolactone)₇]₂₀ (polymer 2). Hydrogen atoms and their corresponding peak integrations are marked by a' and b'.

Table 1Formulation values of independent factors and measured response variables for M-PTX (N = 3: Mean \pm SD).

Run	a	b	c	Response 1		Response 2		Response 3	
	PEGMA-500	CPL	XL	2-h release at pH 6 (%)		2-h release at pH 7.4 (%)		size (nm)	
	No. of units	No. of units	%	Mean (%)	SD	Mean (%)	SD	Mean (%)	SD
1	20	5	25	65.4	2.2	30.9	32.0	21.7	16.9
2	30	6	50	66.3	1.1	39.1	33.9	24.7	20.6
3	30	6	50	70.0	3.1	41.0	34.4	26.1	20.3
4	40	7	25	19.9	2.2	15.7	12.0	9.9	7.0
5	30	7.6	50	112.5	2.2	54.9	55.3	37.5	30.5
6	40	7	75	14.2	2.0	30.4	39.1	23.8	19.4
7	40	5	25	83.1	2.6	36.9	41.5	27.0	21.2
8	46.8	6	50	19.1	1.5	23.5	24.5	16.5	13.0
9	30	6	50	68.7	1.7	40.1	34.6	25.5	20.8
10	30	6	7.9	97.4	2.3	35.9	53.3	30.5	25.9
11	30	4.3	50	106.5	3.3	53.3	51.7	36.1	28.4
12	30	6	50	65.4	1.3	38.9	33.4	24.6	20.3
13	30	6	92	65.4	0.5	52.6	47.1	33.4	28.6
14	20	5	75	96.8	2.6	58.1	49.3	36.7	29.9
15	13.2	6	50	115.5	4.5	56.7	55.8	39.0	29.9
16	40	5	75	60.3	2.5	45.9	38.3	28.9	23.2
17	20	7	75	58.2	1.8	45.0	38.4	28.4	23.3
18	20	7	25	34.0	1.1	20.0	17.0	12.7	10.2
19	30	6	50	67.0	2.0	39.7	33.7	25.1	20.2
20	30	6	50	69.6	2.6	40.7	34.5	25.9	20.5

a, b, c corresponding to number of repeating units in Fig. 1. XL = Crosslinking density was calculated as $c/20 \times 100$, No. of units = Number of repeating units.

platform. Within the DOE domain, we used the central composite design (CCD) to establish a design space with a response surface, using fractional factorial designs with defined center and axial points to estimate the surface curvature. We characterized the synthesized polymers through the shape, size, storage stability and *in vitro* drug release profiles of Cal and PTX in the micelles. Finally, we studied the biodistribution of Cal and PTX from M-Cal/PTX in an aggressive orthotopic mouse model of PDAC with Kras^{G12D} mutation.

2. Materials and methods

2.1. Materials

Cal was purchased from Cayman Chemical Company (Ann Arbor, MI, USA) and PTX from LC laboratories (Woburn, MA, USA). The internal standards (IS) for Cal, calcipotriol-d4 (Cal-d4) and for PTX, paclitaxel-d5 (PTX-d5) were purchased from CRO laboratories Inc. (Dallas, TX, USA). Bead mill homogenizer (Model: Storm BBY24M) and bead lysis kits (Navy RINO screw 10 cap tubes) were purchased from Next Advantage (Troy, NY, USA). LC-MS grade water, methanol, hexane, isopropyl alcohol and dichloromethane were purchased from EMD Millipore Corporation (Billerica, MA, USA). All solutions were ultrasonically degassed before use. Blank C57BL6 whole blood (with Na EDTA) was purchased from Innovative Research (Novi, MI, USA). All other chemical reagents of ACS grade were purchased from Sigma-Aldrich (St. Louis, MO) or VWR (West Chester, PA).

2.2. Polymer synthesis

2.2.1. Polymer reaction initiator, 2-(benzyloxycarbonyl amino) ethyl 2-bromo-isobutyrate (Compound 1)

Cbz-N-ethanolamine 3 g (15.36 mmol) was dissolved in 20 mL of ethyl acetate in a round bottom flask and stirred vigorously with cooling in an ice bath for 1 h. Triethylamine (2.35 mL) was directly added into the mixture in the round bottom flask. 2-Bromoisobutyryl bromide (3 mL, 165.1 mmol) in 5 mL of ethyl acetate was slowly added in dropwise under vigorous stirring. A cloudy product resulted. More ethyl acetate (up to 50 mL) was added when reaction mixture was too viscous. The ethyl acetate solution was subsequently washed with saturated NaHCO₃, 5% HCl, and double-distilled water. Once separated, the aqueous phase (bottom layer) was discarded, and the product was dried with anhydrous

MgSO₄ and condensed in vacuo at 35 °C. The resulting viscous oil was stored at 4 °C (Fig. 1., Compound 1).

2.2.2. Poly[oligo(monomethyl ether ethylene glycol) methacrylate]₄₀-b-poly(2-hydroxyethyl methacrylate)₂₀ (Polymer 1)

The synthetic schemes described in this manuscript are adaptations of the atomic radicalization polymerization reactions described by Matyjaszewski and Xia (Matyjaszewski and Xia, 2002).

Into a round-bottom flask was added Compound 1 (0.17 g, 70.2 μ mol), oligo ethylene glycol methyl methacrylate (OEG₂₀-MA, MW \sim 500, 10 g, 20 mmol), 2,2'-bipyridine, BPy (0.16 g, 1.02 mmol), and 5 mL of anhydrous methanol. The flask was flushed with anhydrous argon for 20 min and immersed in a 55 °C oil bath. CuBr (0.072 g, 0.5 mmol) was quickly added under argon protection, and the reaction was allowed to proceed for 30 min. A separate mixture of 2-hydroxyethyl methacrylate (1.2 mL, 2.1 mmol) in 3.7 mL of de-oxygenized methanol was added into the reaction mixture. The polymerization continued for 18 h at 55 °C and then stopped by exposure to open air for at least 4 h. After a dilution step with ethyl acetate, the reaction mixture was passed through a basic aluminum column to remove the CuBr catalyst and condensed in vacuo at 45 °C to give **Polymer 1** (Fig. 1).

2.2.3. Poly(OEG-MA)₄₀-b-poly[HEMA-g(ϵ -caprolactone)₇]₂₀ (Polymer 2)

Polymer 1 (2.5 g, 0.11 mmol) was dried azeotropically by distillation with toluene at 140 °C and allowed to cool for 1 h. Anhydrous ϵ -caprolactone (CPL, 1.48 mL, 13.14 mmol) and tin (II) 2-ethylhexanoate [Sn(Oct)₂, (0.015 g, 0.04 mmol)] were added. The flask was flushed with anhydrous argon and allowed to run overnight under N₂ gas at 110 °C. The polymerization mixture was precipitated in hexane and kept at -20 °C for 4 h. Once fully precipitated, the product was filtered and dried under N₂ gas for 1 h to obtain 3.8 g of **Polymer 2** (Fig. 1A). The structure of Polymer 2 was confirmed by 1H NMR (Fig. 1B).

2.2.4. Poly(OEG-MA)₄₀-b-poly[HEMA-g(acryloyloxy ϵ -caprolactone)₇]₂₀ (Polymer 3)

To 1 g of **Polymer 2**, 8 mL of tetrahydrofuran (THF) was added, and polymer thoroughly dissolved on an ice bath. To this mixture, 70.5 μ L of triethyl amine (TEA) was added while stirring. To the stirring mixture, 38.3 μ L of acryloyl chloride (0.46 mmol) in 2 mL THF was added dropwise. The mixture reacted for 45 min, then was centrifuged at 6000g

for 10 min after which the supernatant was collected (10 mL) to obtain **Polymer 3** (Fig. 1).

2.2.5. Preparation of micelles loaded with Cal and PTX (M-Cal/PTX) and stability assessments

To 5 mg of PTX and 1 mg of Cal, 2 mL of **Polymer 3** in THF was added and vortexed to ensure the drugs were completely dissolved. The initial drug loading ratio of 5:1 PTX to Cal ratio was chosen after optimization with response surface methodology. Different ratios of PTX and Cal by weight were evaluated with encapsulation efficiency as the readout. We observed a 5:1 ratio of PTX to Cal yielded satisfactory encapsulation efficiencies (>90% for PTX and > 65% for Cal). To the resulting mixture, 10 μ L of tetramethylethylenediamine (TEMED) was added. While vortexing, 4 mL of distilled water was added and vigorously vortexed for 1 min. The organic solvent (THF) was removed in vacuo to leave concentrated micelles (4 mL). Potassium persulfate, K₂S₂O₈ (10 mg) was added and then stirred for 1 h. The loaded micelle (M-Cal/PTX) was dialyzed to remove unencapsulated drugs. Subsequently it was centrifuged at 10,000g for 10 min, and supernatant collected to obtain micelles with a final PTX-to-Cal ratio of ~ 10:1. The freshly prepared micelles were stored at 4 °C for short-term period of 6 weeks. For long-term storage (>3 months), the micelles were stored at -80 °C using 5% sucrose as a cryoprotectant. Stability was evaluated by measuring the changes in micelle size and encapsulation efficiency.

2.3. Micelle optimization using central composite design (CCD)

To obtain micelles with size < 100 nm and extended drug release properties, CCD was employed to tune formulation parameters, namely, the number of repeating units of methacrylate-grafted oligoethylene glycol (OEG-MA-500, a = 13.2 – 46.8 units), CPL (b = 4.3 – 7.6 units), and crosslinking density (XL, 10–90%; c = 2 – 18) (Fig. 1). To simplify the model and reduce the number of parameters to be optimized, the number of repeating units of 2-hydroxyethyl methacrylate was kept constant (20-HEMA) based on preliminary studies. A full factorial design was used to generate models using Design Expert Software v8 (StatEase, Minneapolis, MN) for statistical modeling and the generation of surface response plots. The independent factors of OEG-MA-500, CPL and XL, and their effects on the micelle size, and 2-h accumulative drug release at pH 6 and pH 7.4 were coded at 5 levels as - α , -1, 0, +1, + α , with 0 as the central point (Table 1).

2.4. Characterization of M-Cal/PTX

2.4.1. Cal and PTX encapsulation efficiency

Drug loading efficiency was evaluated by modifying a published protocol (Zhao et al., 2018a). Briefly, micelles were dissolved in methanol and vortexed at high speed, followed by a 5 min sonication to release encapsulated drugs. After centrifugation at 10,000g for 30 min, the supernatant containing released drugs was collected and analyzed on Waters Acquity™ Ultra-high-pressure liquid chromatography (UPLC, Milford, MA, USA). Chromatographic separation was achieved using Kinetex C18 column (1.7 μ m, 100 \times 2.10 mm, Phenomenex, Torrance, CA, USA) under isocratic elution conditions with a total runtime of 3.5 min at a flow rate of 0.4 mL/min. The detection wavelengths for Cal and PTX were 210 nm and 227 nm respectively. The column and autosampler temperatures were 40 °C and 10 °C, respectively, and the injection volume was 10 μ L. The composition of the mobile phase was ammonium acetate solution (5 mM)-methanol (15:85, v/v). Encapsulation efficiency (EE) was computed according to the formula

$$EE(\%) = (\text{Amount of Cal and PTX recovered after dialysis}) / (\text{Amounts Cal and PTX added in micelles}) \times 100\%$$

2.4.2. Morphology and size determination of M-Cal/PTX

Particle size, size distribution, and zeta potential were measured with a dynamic light scattering system on ZetaPlus particle sizer (Brookhaven Instruments Corp., Holtsville, NY). The dried state visualization of particle size and morphology of Cal and PTX-loaded micelles were examined on a transmission electron microscopy (JEOL USA, Inc., Peabody, MA) according to the method described in Zhao et al., 2018b, with digital images (Fig. 4) collected on the AMT Imaging System (Advanced Microscopy Techniques Corp., Danvers, MA).

2.4.3. Drug release of polymeric micelles

To determine the drug release profiles, polymeric micelles containing 1.2 mg/mL PTX and 0.17 mg/mL Cal were diluted in the appropriate matrix (PBS at pH 7.4 or sodium acetate at pH 6, and mouse serum) and added to a microdialyzer (molecular weight cut-off ~ 3500, Thermo Scientific, Rockford, IL). The microdialyzers were incubated in PBS (pH 7.4), sodium acetate buffer (pH 6.0) or mouse plasma in a 37 °C water bath with agitation. Aliquots (30 μ L) were taken at predefined times from the microdialyzer. The samples were centrifuged at 6000g for 5 min and 10 μ L of the supernatant was retrieved and analyzed for Cal and PTX concentrations using UPLC method in Section 2.4.1.

2.5. UPLC-MS/MS assay to measure concentrations of Cal and PTX in biological samples

Separation of Cal and PTX was achieved using the chromatographic conditions described in Section 2.4.1. A triple quadrupole mass spectrometer with electrospray ionization source (ESI) (API 5500, Applied Biosystems/MDS SCIEX, Foster City, CA) was used for the detection using the positive ionization mode with multiple reaction monitoring. Analyte extraction from biological samples was achieved with a liquid-liquid extraction procedure. Whole blood samples were processed using a developed and validated UPLC-MS/MS assay for simultaneous quantifications of Cal and PTX (In press; Lincha et al., 2020).

Tissues samples were weighed and homogenized with bead mill homogenizer in 1 mL of water at 4 °C. The tissue homogenate was transferred into a clean 1.5 mL Eppendorf tube from which 100 μ L was extracted with 500 μ L of water-MeOH (1:1, v/v) containing IS (10 ng/mL each of PTX-d5 and Cal-d4) by vortexing for 1 min. Additional processing and analyte measurement were as described in the referenced manuscript. Concentrations of analytes in tissues were normalized by the tissue weight.

The UPLC-MS/MS assay was linear over a range of 0.5 – 500 ng/mL for both Cal and PTX with the lower limits of quantifications (LLOQs) of 0.5 ng/mL in whole blood, tumor, liver and spleen.

2.6. Biodistribution studies

All animal studies were approved by the Institutional Animal Care and Use Committee (IACUC) and were conducted in accordance with institutional guidelines. Kras* murine pancreatic cancer cells with a doxycycline-inducible mutation of KRAS^{G12D} were cultured and the Kras* murine PDAC model was developed according to a previously published protocol (Zhao et al., 2018b). Eight-weeks old C57BL/6 female mice (Taconic Biosciences, Rensselaer, NY) were injected with Kras* cells into the pancreas head under isoflurane anesthesia after a small opening was made in the abdomen to expose the pancreas. A 2-

Table 2

Equations for best-fit models and summary statistics for micelle size and 2-h PTX release at pH 6 and 7.4.

Response variables	Model equations in terms of coded factors	R ²	p value
2-h PTX release at pH 6	$47.49 - 19.77 \times \text{PEGMA-500} + 16.83 \times \text{CPL}$	0.62	0.0035
2-h PTX release at pH 7.4	$48.34 - 19.70 \times \text{PEGMA-500} + 11.19 \times \text{CPL} + 2.97 \times \text{XL} - 13.96 \times \text{PEGMA-500} \times \text{CPL} + 11.26 \times \text{CPL}^2 - 8.26 \times \text{XL}^2$	0.83	0.0004
Micelle size	$1.72 - 0.13 \times \text{PEGMA-500} - 0.11 \times \text{CPL} + 0.10 \times \text{XL}$	0.60	0.0262

gauge needle was used for precise injection of 5×10^5 Kras* cells. Mice with palpable pancreatic tumors of diameter 5 mm were recruited and randomized into two study groups. The mice were each administered an intravenous bolus of M-Cal/PTX (N = 3) at doses of 5 mg/kg PTX and 0.5 mg/kg Cal or the non-formulated drugs (N = 3) at the equivalent doses. Blood samples were collected at pre-determined time points and the mice were euthanized 24 h post dose. Tumor and liver tissues were collected for simultaneous quantifications of Cal and PTX concentrations using the developed and validated tandem UPLC-MS/MS assay referenced in Section 2.5.

2.7. Statistical analysis

Data were evaluated using Student's *t*-test or 1-way analysis of variance (ANOVA) followed by post hoc Tukey for multiple comparisons. The statistical significance was evaluated at $p < 0.05$.

3. Results and discussion

3.1. Development of an optimal M-Cal/PTX using CCD

3.1.1. Selection of an optimal formulation with PTX as the model drug

The effects of the number of units of OEG-MA-500, CPL, and XL (%) on the size and 2-h drug release kinetics of PTX were studied using 20 experimental runs (Table 1). PTX was chosen as the model drug for CCD formulation optimization, because PTX (MW = 853.9 g/mol, logP = 3.52, water solubility = 0.0056 mg/mL) and Cal (MW = 412.6 g/mol, logP = 4.3, water solubility = 0.0135 mg/mL) have similar physico-chemical properties, making extrapolation from PTX to Cal feasible. Economic consideration was another factor for not using large amounts of Cal in the process of formulation optimization but validating with Cal in the optimized formulation. Micelle size from the experimental runs ranged between 16.6 and 100 nm, cumulative 2-h PTX release at pH 6 and 7.4 were 17.7–115.4% and 6.6–100%, respectively, depending on the formulation compositions. The formulation variable-dependent drug release characteristics were best described by a linear model for pH 6

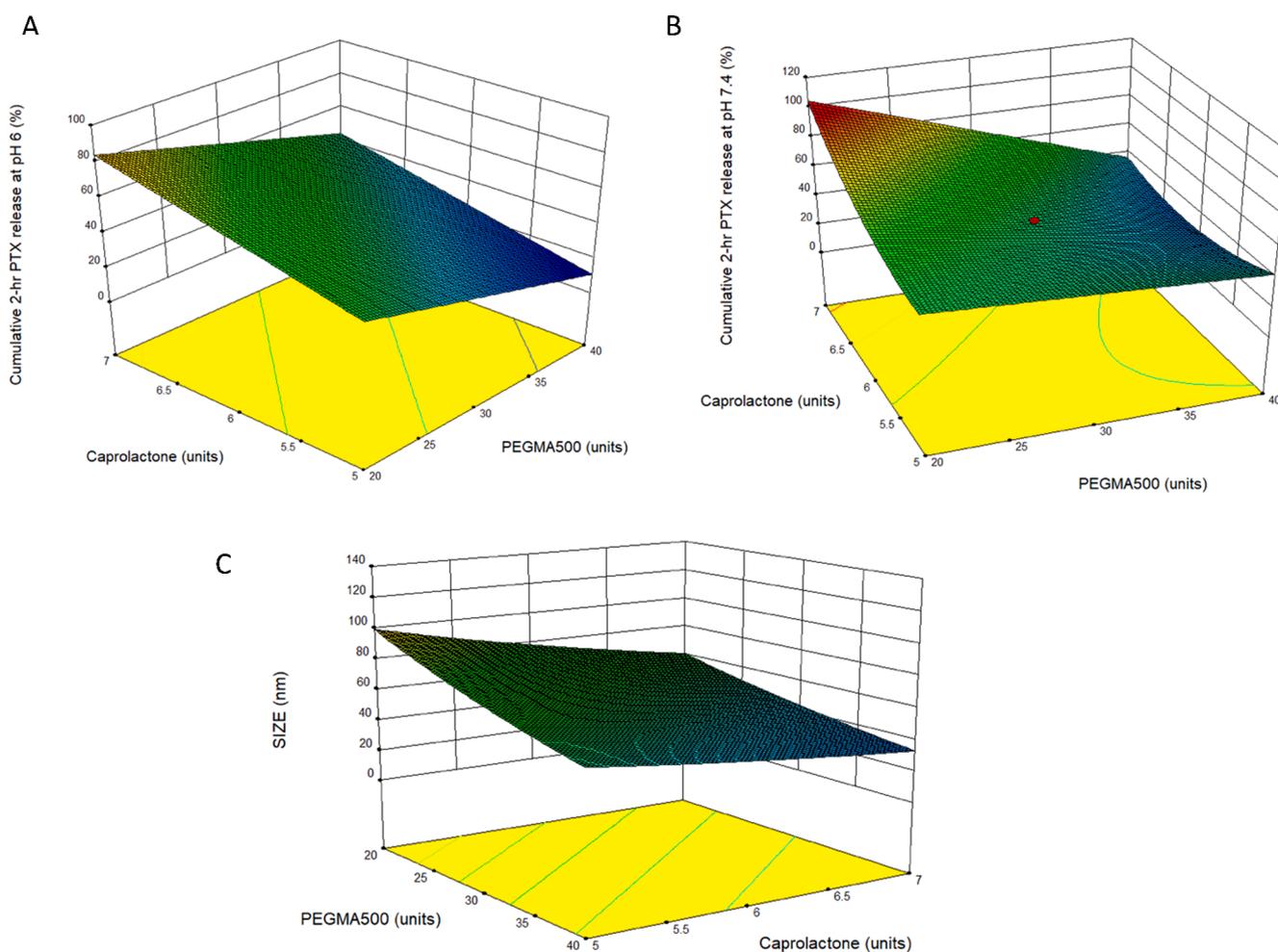


Fig. 2. Three-dimensional response surface plots showing the effects of polymer variables of PEGMA-500 and CPL units on A (PTX release at pH 6), B (PTX release at pH 7.4) and C (micelle size). For graphing purposes, the crosslinking density was set at 50%

and, and a quadratic model for the release characteristics for pH 7.4. A log-transformed linear model best described the impacts of formulation variables on the micelle size, as summarized in Table 2. Positive values of regression were indicative of synergism between the dependent and independent variables, while negative coefficients signified antagonism (Hao et al., 2012). Surface response maps corroborated the mathematical models and showed that the extent of drug release decreased with increasing amount of PEGMA-500 at pH 6. However, at pH 7.4 the opposite trend was observed (Fig. 2A and B), while micelle size also decreased with higher PEGMA-500 amount (Fig. 2C). The decrease of nanoparticle size with increasing PEGMA-500, a capping agent, was anticipated, as it has long been recognized and discussed (Arulmozhi and Mythili, 2013). PEGMA-500, a long chain polymer with terminal hydroxy groups caps polymer growth by shielding and stabilizing the nanoparticle. As more PEGMA-500 is added, a greater amount of OEG groups are present to cap the end of particles, effectively reducing average size (Arulmozhi and Mythili, 2013). Since the first report of PEGylation for drug delivery (Abuchowski et al., 1977), this strategy has become a mainstay in drug formulation due to the unique stealth properties, resisting interaction with components of a biological matrix, usually blood that it confers on drugs (Suk et al., 2016). It is therefore no surprise that as the amount of PEGMA-500 increases, the micelle stabilization increases, and the cumulative PTX release decreases. This mechanism of extending circulation times of drugs underlines the reason why PEGylation is widely used to formulate drugs when longer circulation is desired (Arturson et al., 1983; Tan et al., 1993).

Unlike PEGMA-500, as CPL increased, the cumulative release of PTX over 72 h increased. The micelle size decreased with increasing CPL, with a negative correlation. CPL has widespread applications in drug delivery for its biocompatibility, biodegradability and being generally recognized as safe (Zelenková et al., 2014). Additionally, CPL undergoes slow degradation in the body (Karuppuswamy et al., 2015; Seremeta et al., 2013; Woodruff and Hutmacher, 2010). These properties make CPL suitable for drug delivery. It is widely expected to slow down drug release with increasing CPL amount, due to increased hydrophobic interaction between the polymeric matrix and the drug payloads. However, our observation was the opposite. We speculated that the interaction between CPL and other polymers might be the reason. CPL has excellent compatibility with other polymers in the formulation, including PEG, and therefore it is plausible the characteristics of this block of copolymer may change as it interacts with other ingredients. The compatibility of CPL with PEG makes it desirable when it is intended to control properties like degradation kinetics and hydrophilicity (Bilensoy et al., 2009; Payyappilly et al., 2015). The interaction between PEGMA-500 and CPL was indeed observed in our model to affect the PTX drug release at pH 7.4. This interaction was captured by the curvature for the surface response map from our CCD approach (Fig. 2B), and the mathematical model suggested the impact of the interaction was dominated by PEGMA-500.

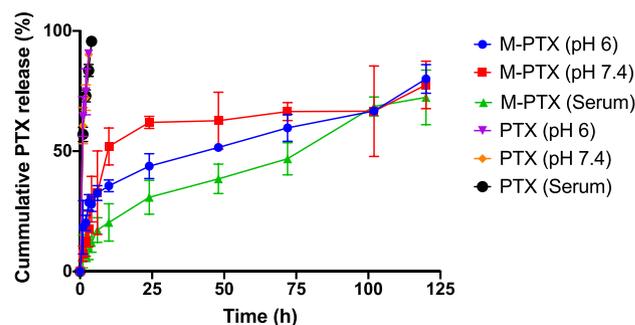
To further improve stability of the polymeric micelles, we extended the end units of CPL with acryloyloxy to form crosslinks in the hydrophobic domain of the block copolymer by potassium persulfate mediated reaction. The goal was to obtain micelles that released drugs while in circulation but remained stable enough to control release. We therefore studied the extent of XL on the size and drug release characteristics. The XL did not significantly affect the drug release at pH 6, and thus was dropped and set at 50% for model simplicity and visualization efficiency (Table 2), in subsequent studies to generate the surface response maps. The optimal micelle consisted of 40 PEGMA-500 repeating units ($a = 40$), 7 repeating units of CPL ($b = 7$), 20 units of HEMA, and 75% crosslinking density (corresponding to $c = 15$). The structure of Polymer 2 from which polymer 3 and the optimal micelles were formed in one pot is confirmed by ^1H NMR (Fig. 1B).

Table 3

Model-predicted and observed outputs for PTX in optimal M-Cal/PTX.

Optimal M-Cal/PTX	Predicted	Observed N = 3 Mean + SD)	%Bias
PTX size (nm)	53	51 ± 2.7	3.7
2-h Cumulative release (%)			
pH 6	25	20 ± 4.5	20
Ph 7.4	22	18 ± 3.3	18.2

A.



B.

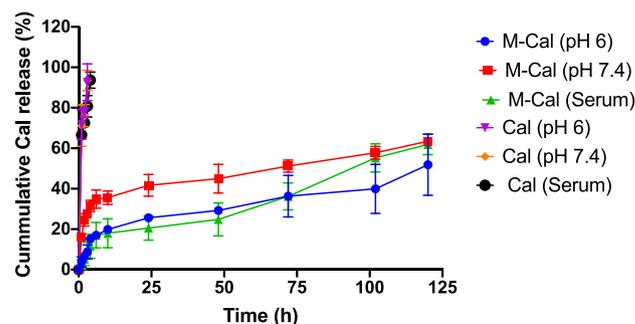


Fig. 3. Drug release profiles of Cal and PTX from optimal M-Cal/PTX (M-Cal, M-PTX) and from a 10% dimethyl acetamide cosolvent (Cal, PTX). Graph shows mean \pm SD of 3 independent studies and an insert for 0–4 h.

3.1.2. Selection of an optimal drug delivery system and validation of the CCD with Cal and PTX

Numerical optimization was used to obtain the optimal drug-loaded micelles. A desirability function was used to simultaneously optimize response by indicating a range for each independent variable within the experimental design. The limits for each response was assigned a minimum, maximum or target value. To allow flexibility in model predictions, a range for each response was indicated. The objectives of developing this drug delivery system were to minimize Cal toxicity by reducing free Cal in systemic circulation, extend the apparent biological half-life of Cal and PTX, and enhance their accumulation in the tumor. Based on preliminary experiments, we aimed at having no more than 25% of drugs released at 2 h to control initial burst effect, and micelle size range of 40–100 nm. The range for XL was set at 50–75%. Even though PEGMA-500 confers stealth properties on nanoparticles and increases circulation time, its hydrophilicity reduces cellular uptake (Ruiz et al., 2013; Wang and Thanou, 2010). However, nanoparticle size and shape also strongly impact cellular uptake (Albanese et al., 2012; Zhang et al., 2015). We desired micelles with a size of 40–100 nm, because this is the optimal size for cellular uptake (Jiang et al., 2008; Lu et al., 2009; Yuan et al., 2010; Zhang et al., 2009). Nanoparticles within such a size range have the appropriate entropic and enthalpic properties that influence adhesion strength between the particles and cellular receptors (Yuan et al., 2010; Zhang et al., 2009). The optimal micelle was thus

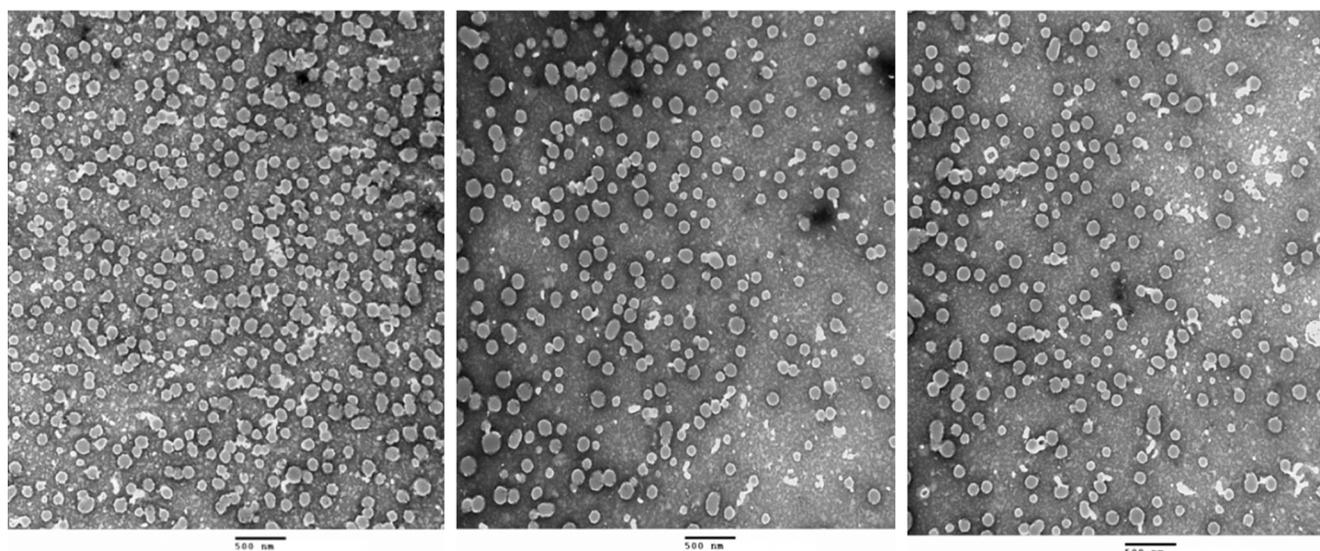


Fig. 4. TEM images of M-Cal/PTX at a scale of 500 nm and at 25,000X magnification (Images shown were from three sets of independently prepared M-Cal/PTX).

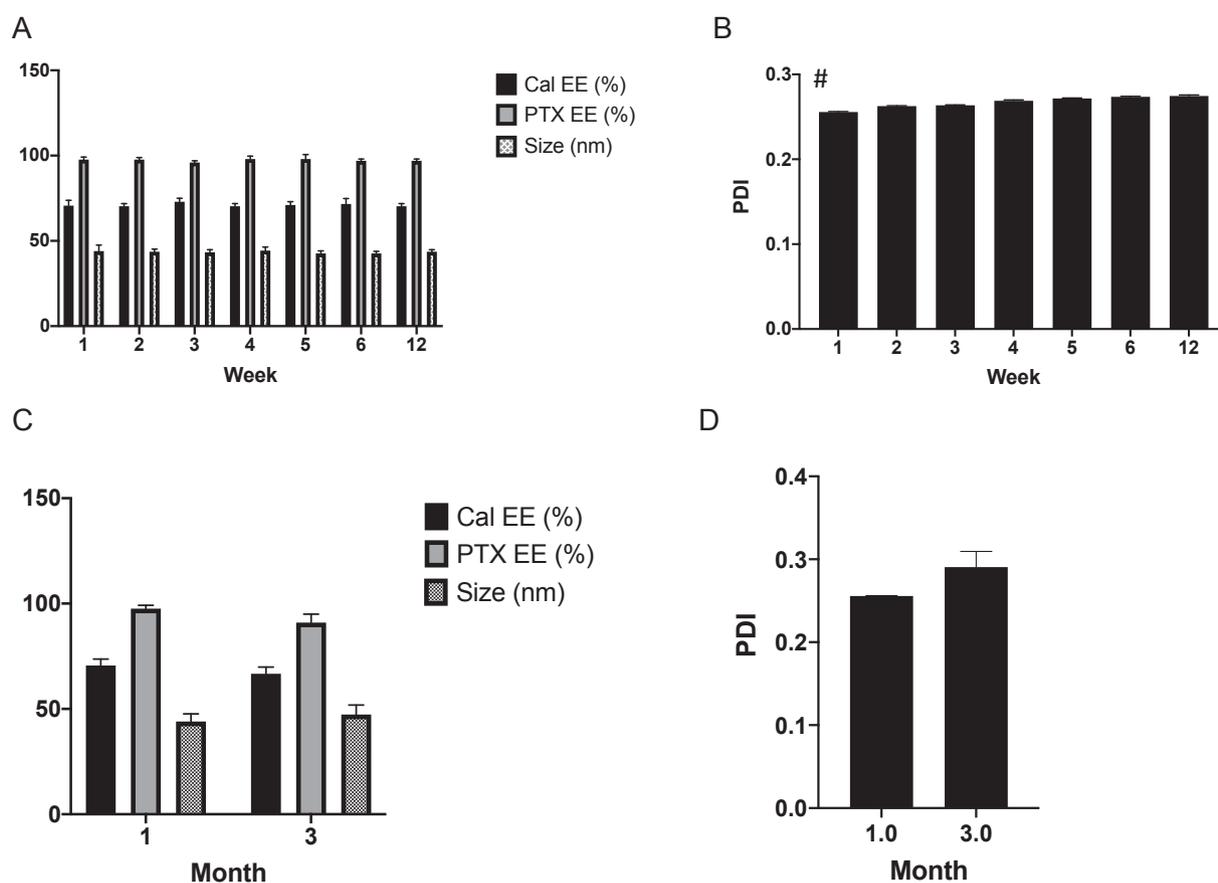


Fig. 5. Storage stability of optimal M-Cal/PTX at 4 °C (A, B) and -80 °C (C, D). Data shows mean \pm SD of measurements from 3 independently prepared batches. * $p < 0.05$ ($n = 3$).

selected with 40 repeating units of PEGMA-500 ($a = 40$), 20 repeating units of HEMA and 6 repeating units of CPL ($b = 6$) along with 75% XL ($c = 15$). The predicted average values of particle size, 2-h drug release at pH 6 and 7.4 were 53 nm, 25% and 22%, respectively. The model was validated by performing confirmatory runs using the model-predicted optimal micelle composition, with co-encapsulation of Cal and PTX. The observed values for average micelle size was 51 nm. For PTX, 2-hour

drug release at pH 6, 7.4 and in serum were 20%, 18% and 7%, respectively, and for Cal, 6%, 24% and 6% (Table 3). We postulated that the initial slow release of Cal and PTX in serum when compared to buffers tested could be due to binding of released drugs to serum. The observed responses correlated well with model-predicted response with biases between 3.7% and 20% highlighting the reliability of using the CCD model for formulation development (Table 3). The full drug release

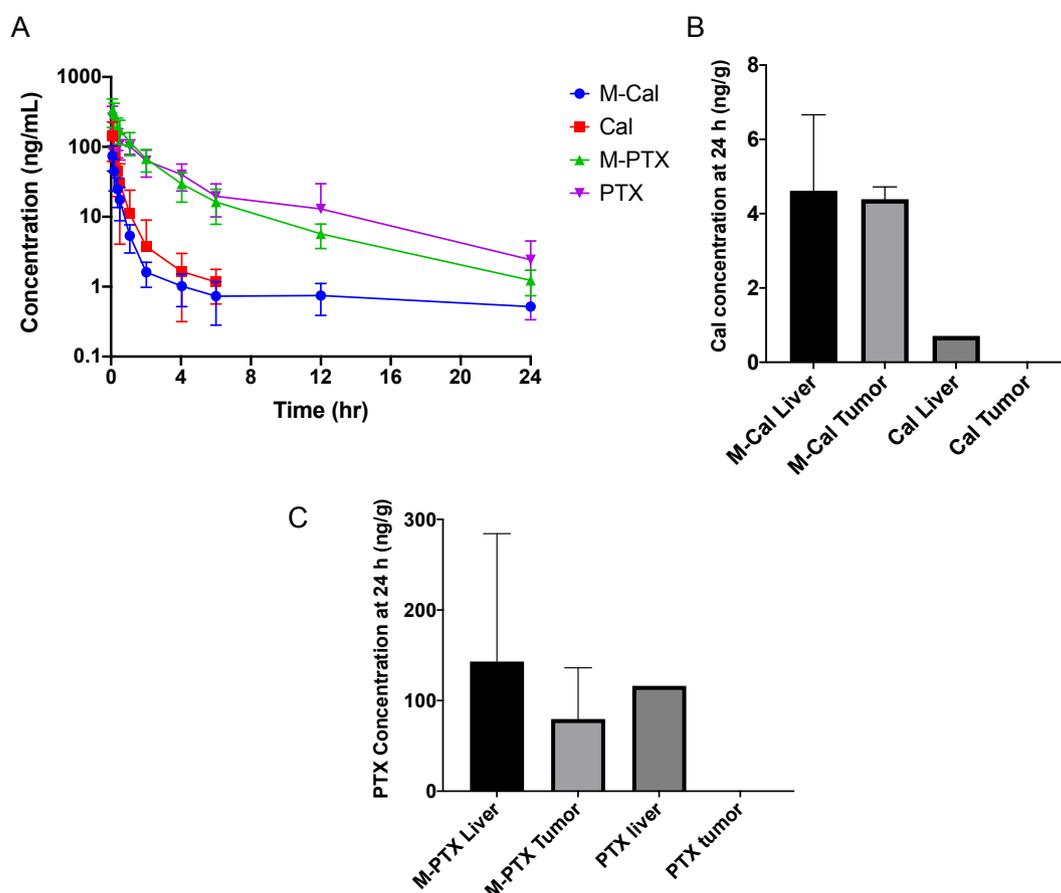


Fig. 6. Pooled whole blood PK profiles (A) and tissue biodistribution of optimal Cal (B) and PTX (C) from M-Cal/PTX and unformulated Cal/PTX at 24 h. Mice were administered a single IV bolus dose of 5 mg/kg PTX and 0.5 mg/kg Cal. Free Cal/PTX was prepared in 10% dimethylacetamide. Concentration time profiles were constructed from N = 8 for M-Cal/PTX and N = 7 for Cal/PTX groups. For the biodistribution study data show N = 3 for M-Cal/PTX and free drugs groups. For mice received free Cal and free PTX, the drugs were measurable in 1 out of the 3 animals in the liver, and drug concentrations were below detection limit in the tumor in all animals. For Fig. 6A, free Cal and PTX were not measurable beyond 4 and 12 h respectively. Data are presented as mean \pm SD.

profile (Fig. 3) showed that the cumulative percentage of Cal and PTX released at pH 6, 7.4 and in serum were 42%, 58%, 58% and 80%, 80%, 76%, respectively after 120 h, demonstrating the sustained drug release characteristics of the optimal micelles.

3.2. Characterization of optimal M-Cal/PTX

3.2.1. TEM analysis

TEM analysis of the optimal M-Cal/PTX showed spherical particles with the size range of 40–100 nm (Fig. 4). The shape and size distribution were consistent among the three independently prepared micelles.

3.2.2. Encapsulation efficiency of optimal M-Cal/PTX

The initial ratio of PTX to Cal (w/w) affected the encapsulation efficiencies of both drugs. To obtain satisfactory encapsulation efficiencies, a DOE was used to evaluate different combinations of both drugs with encapsulation efficiency as the readout. A drug loading ratio of 5:1 PTX to Cal yielded a high PTX encapsulation efficiency of > 90% and an encapsulation efficiency of > 65% for Cal.

3.2.3. Polydispersity index and zeta potential of optimal M-Cal/PTX

The size distribution of the optimized formulation, reflected by PDI measured by dynamic light scattering (DLS), was 0.249 ± 0.004 and a zeta potential of 0.006 ± 0.001 (N = 3 independent batches). In general, particles with a PDI < 0.3 are considered uniformly dispersed (Das and Chaudhury, 2011)

Table 4

Whole blood PK parameters of Cal and PTX from M-Cal/PTX or unformulated Cal/PTX in healthy C57BL/6 mice after a single IV bolus dose of 5 mg/kg PTX and 0.5 mg/kg Cal, N = 3 in each group.

		M-Cal		Cal		M-PTX		PTX	
$t_{1/2}$	h	3.3 [#]	(1.8)	0.9	(0.2)	4.5	(1.1)	3.4	(0.4)
C_{max}/D	(ng/mL)/(mg/kg)	124.5	(29.3)	315.5	(178.6)	70.3	(29.4)	60.2	(26.7)
AUC_{last}/D	$h^*(ng/mL)/(mg/kg)$	71.6	(22.6)	166.1	(117.2)	106.4	(30.4)	123.2	(41.4)
AUC_{INF}/D	$h^*(ng/mL)/(mg/kg)$	79.3	(29.9)	170.4	(120.7)	108.3	(30.2)	127.1	(44.3)
CL	L/h/kg	14.0	(4.7)	7.6	(3.5)	9.9	(3.3)	8.7	(3.1)
MRT_{inf}	h	1.8	(1.3)	0.7	(0.08)	3.5	(0.8)	3.6	(0.7)
V_{ss}	L/kg	22.2 [#]	(10.5)	5.5	(2.30)	33.2	(8.8)	36.4	(7.2)

Values shown are Mean (SD). [#] Parameter is significantly different from free Cal group, by Student's *t*-test at $p < 0.05$.

3.2.4. Storage stability at 4 °C and -80 °C

The stability of M-Cal/PTX in storage is shown in Fig. 5. Particle size and EE (Fig. 5A), and PDI (Fig. 5B) were measured over a 6-week period from micelles stored at 4 °C. The micelle size and encapsulation efficiencies of both drugs were not appreciably changed over the 6-week period and also after 3 months when stored at -80 °C in 5% sucrose. The PDI increased from 0.25 to 0.26 after 1 week to 0.27 after 6-week storage; nevertheless, but was still below the stable threshold PDI of 0.3. The optimized M-Cal/PTX formulation had a 75% crosslinking density and could be used within 6 weeks of storage at 4 °C after preparation. For long term storage, micelle integrity is maintained by storing at -80 °C in 5% sucrose up to 3 months. We attributed the stability of the micelles to the effective core crosslinking. The end hydroxy units of HEMA were conjugated with acryloyl chloride to enable crosslinking in the presence of potassium persulfate.

3.2.5. Pharmacokinetic and biodistribution of optimal M-Cal/PTX in healthy mice and an orthotopic Kras^{G12D} mouse model of pancreatic cancer

The pharmacokinetic (PK) profiles of Cal (from dosing free Cal, dissolved in 10% dimethyl acetamide), PTX (from free PTX, dissolved in 10% dimethyl acetamide), M-Cal, and M-PTX (from dosing M-Cal/PTX) in healthy C57BL/6 mice were presented in Fig. 6A. The PTX and M-PTX profiles appeared similar with no statistical difference at any time point. However, while M-Cal could be measured up to 12 h post dose, Cal could only be measured up to 4 h post dose. In addition, the elimination phase of free Cal appeared steeper than that of Cal in M-Cal. The biodistribution of Cal (Fig. 6B) and PTX (Fig. 6C) in tumor of PDAC Kras* tumor from M-Cal/PTX at 24 h post dose were clearly higher than those from free Cal and free PTX. The Cal in liver from dosing of M-Cal/PTX was higher than that from free Cal (Fig. 6B), but those of PTX were similar between M-Cal/PTX and free PTX (Fig. 6C). The blood PK parameters were estimated using Phoenix® version 8 software (Table 4). One of the main goals for developing the micellar formulation was to reduce systemic exposure of Cal. Predictably, the dose-normalized AUC_{inf}, AUC_{last} and C_{max} of Cal was lower in mice administered the M-Cal/PTX, when compared to those that received the free Cal (Table 4). Specifically, the dose normalized AUC_{inf}, AUC_{last} and C_{max} of micelle-treated group appeared to be >2 times lower than in free drug group, albeit not statistically significant. However, the corresponding parameters for PTX in both groups were relatively similar. We postulated that off-target activation of VDR is directly or indirectly responsible for some of the toxic side effects associated with Cal. Therefore, it was our expectation that reduced Cal exposure would correlate with the potential reduced toxicity of Cal (Table 4).

With the dose of M-Cal/PTX, the apparent elimination half-life (t_{1/2}) of Cal was 3.7 times longer than that with free Cal. The prolonged half-life resulted from the increased V_{ss}, suggesting a greater particle uptake and longer residence time in tissues from M-Cal/PTX, and also consistent with the extended drug release characteristics of the micellar formulation. On the other hand, the t_{1/2} of PTX was not appreciably prolonged with M-Cal/PTX formulation.

We also measured Cal concentrations in liver and tumor from unformulated Cal or PTX and M-Cal/PTX. After a single IV bolus dose, only M-Cal/PTX yielded sustained levels of drugs in the tumors (Fig. 6B and 6C). These results confirmed the micellar drug delivery platform facilitated tumor accumulation. The shape and size are two physical characteristics of nanoparticles that play crucial roles in tumor uptake (Batist, 2007). Nanoparticles face several biobarriers while in circulation. A major barrier to tumor uptake is the body's immune response which considers the formulation platform a foreign body. However, tuning the particle size and shape can provide stealth allowing the particles to circulate longer and accumulate in desired areas through the "Enhanced Permeability and Retention" (EPR) effect (Batist, 2007; Lasic and Papahadjopoulos, 1995). Nanoparticles of size more than 200 nm do not generally extravasate into tumor (Nagayasu et al., 1999). Also, shapes of nanoparticles dictate their interactions with membranes and

circular shapes are favored for tumor accumulation (Nagayasu et al., 1999). A combination of these factors possibly contributed to the enhanced uptake of Cal and PTX encapsulated in the M-Cal/PTX system.

4. Conclusion

Herein, we developed a micellar drug delivery platform to encapsulate and deliver Cal and PTX simultaneously for the treatment of pancreatic cancer. A sustained release delivery system was desired to reduce the systemic exposure of Cal in the circulation, a potential solution to Cal-associated toxicity. Additionally, micelles with a particle size < 100 nm were desired to improve tumor accumulation. We used the DOE approach to obtain the optimal delivery system with the size range of 40–100 nm which resulted in longer apparent biological half-life of Cal. The encapsulation efficiencies of Cal and PTX were sufficient to deliver therapeutically relevant doses in a mouse model of PDAC. Biodistribution studies using the micellar drug delivery system yielded Cal and PTX accumulation in tumors 24 h after a single IV bolus, which was not replicated in mice that received non-encapsulated Cal and PTX. We postulated that the sustained release characteristics, size and shape of the optimal formulation permitted the sustained tumor accumulation. On-going studies will focus on demonstrating the proof-of-concept efficacy and safety of this micellar drug delivery system in relevant mouse models of PDAC.

CRedit authorship contribution statement

Victor R. Lincha: Conceptualization, Investigation, Formal analysis, Methodology, Writing - original draft, Visualization. **Jun Zhao:** Conceptualization, Investigation, Formal analysis, Project administration, Supervision. **Xiaoxia Wen:** Investigation, Methodology. **Chiyi Xiong:** Supervision. **Diana S-L Chow:** Conceptualization, Funding acquisition, Project administration, Resources, Validation. **Chun Li:** Conceptualization, Funding acquisition, Project administration, Resources, Validation.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ijpharm.2021.120523>.

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